SYNTHESIS OF CARBON-14 LABELLED ACETAMINOPHEN (4-HYDROXYPHENYLACETAMIDE, CARBONYL-14C)

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A simple and convenient method for the condensation of paminophenol (I) with ^{14}C -acetic anhydride (II) yields up to 76% of ^{14}C -labelled 4-hydroxyphenylacetamide (carbonyl- ^{14}C) (III). Typical synthesis time is 60 minutes. Radioactive yield is 51%, and specific activity is 0.22 $\mu\text{Ci/mg}$.

Key Words: 14C-hydroxyphenylacetamide (carbonyl-14C), 14C-acetaminophen

REACTION

INTRODUCTION

Application of labelled compounds is still a sensitive and selective tool for metabolic and pharmacokinetic studies. Although ³H-acetaminophen is available commercially, the inherent impurity, which is described (1), restricts its usefulness. Although a purification method is available, it is time consuming.

Numerous synthetic methods for acetaminophen have been reported in the literature (2-7). However improvements are needed in purity, yield and synthetic time.

We now present a simple and convenient method for the synthesis and purification of $^{14}\mathrm{C}\text{-acetaminophen}$.

EXPERIMENTAL

Purification of p-aminophenol

In a 250 ml beaker, 21.8 g of p-aminophenol was dissolved in 130 ml of deionized water at 90°. To this solution 0.04 g of sodium dithionite was added as antioxidant. Finally, 1 g of Fe-free charcoal was added. The resulting solution was heated to boiling for five minutes, filtered and cooled to yield p-aminophenol. This procedure was repeated several times until the product was colorless.

Preparation of Fe-free charcoal

A small amount of citric acid (approximately 1 g) was dissolved in a dilute solution of hydrochloric acid. This solution was then used to wash the charcoal in order to eliminate any Fe present.

Synthesis of 4-hydroxyphenylacetamide

A "break seal" glass ampoule containing 1 mCi of 14 C-acetic anhydride b (specific activity 28.6 mCi/m mole), filtered with a reflux condenser, was cooled (dry ice bath), the seal broken and 0.94 ml of acetic anhydride, 1.09 g of dry, freshly purified p-aminophenol and 10-15 drops of deionized water was added. The reaction mixture was stirred and heated to 70° for 20 minutes and cooled. The 14 C-labelled 4-hydroxyphenylacetamide which precipitates was filtered and recrystallized from deionized water with Fe-free charcoal to yield white crystals (mp 166-169°, chemical yield 76%, specific activity 0.22 μ Ci/mg, radioactive yield 51%).

Radioactive purity of the final product was determined by thin layer chromatography using silica gel with fluorescent indicator^C and 2-butanone as

^a Aldrich Chemical Co., Milwaukee, Wisconsin, USA

b Amersham Searle Co., Arlington Heights, Illinois, USA C Silica gel GF, Analtech Inc., Newark, Delaware, USA

mobile phase. Only one spot was observed under exposure to short UV light. This separation was characterized by an $R_{\mathbf{f}}$ value of 0.55. In addition, two dimensional ascending chromatography was performed using chloroform-acetone-cyclohexane (50:30:20), followed by benzene-ether-methanol acetic acid (60: 30:5:5) as mobile phases (1). These solvent systems gave separation characterized by $R_{\mathbf{f}}$ values of 0.2 and 0.3, respectively. Only one spot was observed under exposure to short UV light. Radioactive scanning of the plate showed over 95% purity. No radioactive impurities were detected on the tlc plates.

Structure determination was confirmed by infrared, nmr and ultra-violet spectroscopy.

All radioactivity measurements were made using a Packard Tri Carb Scintillation Spectrometer, Model 3375, with efficiency of 90% and background count of 30 cpm. The samples were counted in a liquid scintillation cocktail containing 30.3 g of 2-(4'-t-butylphenyl)-5-(4"-biphenyl)-1,3,4-oxidiazole and 1.9 g of 2-(4'-biphenyl-6-phenyl)-6-phenylbenzoxazole made up to a gallon with freshly distilled toluene.

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